Use of Protein Cross-Linking and Radiolytic Labeling To Elucidate the Structure of PsbO within Higher-Plant Photosystem II

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ABSTRACT: We have used protein cross-linking with the zero-length cross-linker 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide, and radiolytic footprinting coupled with high-resolution tandem mass spectrometry, to examine the structure of higher-plant PsbO when it is bound to Photosystem II. Twenty intramolecular cross-linked residue pairs were identified. On the basis of this cross-linking data, spinach PsbO was modeled using the Thermosynechoccus vulcanus PsbO structure as a template, with the cross-linking distance constraints incorporated using the MODELLER program. Our model of higher-plant PsbO identifies several differences between the spinach and cyanobacterial proteins. The N-terminal region is particularly interesting, as this region has been suggested to be important for oxygen evolution and for the specific binding of PsbO to Photosystem II. Additionally, using radiolytic mapping, we have identified regions on spinach PsbO that are shielded from the bulk solvent. These domains may represent regions on PsbO that interact with other components, as yet unidentified, of the photosystem.

Photosystem II (PS II) is a light energy-driven water-plastoquinone oxidoreductase. Excitation energy from the light-harvesting apparatus (phycobilisomes in cyanobacteria and light-harvesting chlorophyll proteins in green plants and algae) is transferred to the reaction center of the photosystem. Initial charge separation occurs between ChlD1 and PheoD1, which yields ChlD1•−PheoD1•+.

This charge-separated state is stabilized by rapid electron transfer from PheoD1•+, first to QA and then to QB. The accumulation of two electrons on QB leads to protonation and its subsequent release as plastoquinol. ChlD1•− is reduced by P0D1• and PheoD1•+ is reduced by Yz, the residue D1:161Y, yielding Yz• with the release of a proton. The subsequent reduction of Yz• by the Mn4CaO5 cluster occurs via proton-coupled electron transport and leads to the accumulation of an oxidizing equivalent at the oxygen-evolving site, the Mn4CaO5 metal center. The accumulation of four oxidizing equivalents ultimately leads to the oxidation of two water molecules and the release of dioxygen.2,3

Cyanobacterial PS II contains at least 20 subunits, of which at least 17 are intrinsic membrane proteins.4 A subset of these components (D1, D2, CP43, CP47, the α and β subunits of cytochrome b559, and the PSBL protein) is required for oxygen evolution and accumulation of the photosystem in thylakoid membranes. Genetic or biochemical removal of any of these protein subunits leads to loss of functional PS II. In addition, 10 low-molecular mass intrinsic proteins are present; however, their functions remain obscure. PS II complexes containing only these intrinsic proteins evolve oxygen at low rates and require very high concentrations of calcium and chloride.5,6 At presumptive physiological concentrations of these cofactors, efficient oxygen evolution requires the extrinsic components of the photosystem. These include PsbO, which is uniformly present in all oxygenic organisms, and either PsbU, PsbV, CyanoQ and CyanoP (in the cyanobacteria), or PsbP, PsbQ, and PsbR (in green plants) (for reviews, see refs 7–12). PsbO plays an important role in the stabilization of the Mn4CaO5 cluster13 at low chloride concentrations and protects the metal cluster from exogenous reductants. In higher plants, it is required for PS II assembly7,14–17 and photosynthetic oxygen evolution. Interestingly, in cyanobacteria, this is not the case.17 In the absence of PsbO, Synechocystis can grow autotrophically and evolve oxygen, albeit at lower rates.18

Over the past 15 years, increasingly higher resolution crystal structures of thermophilic cyanobacterial PS II have become available.4,19–24 These studies have been critically important in improving our understanding of the molecular organization of the photosystem. Recently, a crystal structure of PS II from the red alga Cyanidium caldarium, at 2.76 Å resolution, has also become available.25 This is, importantly, the first structure of PS II available from a eukaryote. It should be noted, however, that the red algae do not lie in the green plant lineage.

As no crystal structure for higher-plant PS II is available, the structure and interactions of the extrinsic proteins with the
intrinsic components of the photosystem have been modeled by comparison to those of cyanobacterial PS II. It must be stressed, however, that the interactions of the PsbP and PsbQ proteins with PS II are not well understood. Additionally, strong biochemical evidence indicates the presence of a putative second copy of PsbO being associated with each PS II monomer in higher plants (for reviews, see refs 13 and 26). Protein cross-linking coupled with tandem mass spectrometry is a powerful technique for elucidating protein–protein interactions in multisubunit protein complexes.27,28 Recently, these methods have been used to investigate subunit–subunit interactions within PS II and PS I.29−32 Liu et al.30 elucidated the structure of a PS II–phycobilisome–PS I megacomplex using dithiobis (succinimidylpropioniate). Liu et al.31 using the cross-linkers 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC, a zero-length cross-linker) and dithiobis(succinimidylpropionate), identified a binding domain for CyanoQ within Synchocystis PS II. Ido et al.32 have extended their earlier studies using the cross-linker EDC on higher-plant PS II. In this study, they identified interchain cross-linked products involving PsbP, PsbQ, and other PS II components (CP43, CP26, and PsbR). Ido et al.32 have positioned PsbQ on the periphery of the PS II complex in marked contrast to the position of CyanoQ proposed by Liu et al.31 Our laboratory has used the cross-linker bis-sulfosuccinimidyl suberate to examine the interaction of PsbP and PsbQ in higher-plant PS II.33 Our results indicate that PsbP and PsbQ directly interact, with the location of PsbQ being consistent with the location hypothesized by either Liu et al.31 or Ido et al.32 It should be noted that the proposed location of PsbQ presented by Ido et al.32 and modified by Mummadisetti et al.33 is similar to the location of PsbQ in the Cyanidium PS II crystal structure.34 Recently, models for the structural organization of the higher-plant extrinsic proteins in association with PS II have been presented.12

The radiolysis of water by synchrotron radiation produces \( ^{1}O_{2} \), which is capable of oxidatively modifying amino acid residues that are in contact with the bulk solvent. The modified residues can be identified using tandem mass spectrometry, and their location can be used to "footprint" protein–protein interactions within multisubunit protein complexes.35−36 This radiolytic mapping technique has been used to identify both surface and buried amino acid residues that are exposed to water.37−40 Recently, we used this technique to examine the interaction of PsbP and PsbQ in higher-plant PS II.33 In this study, we identified domains on PsbP that appear to be shielded from the bulk solvent by intrinsic components of PS II and, possibly, by PsbR.

In this paper, we have used protein cross-linking coupled with tandem mass spectrometry to identify 20 intramolecular cross-linked products within the PsbO protein. These experiments allowed the modeling of the N-terminus of higher-plant PsbO, which contains a 10-amino acid extension when compared to cyanobacterial PsbO. This facilitated the localization of the two binding determinants for higher-plant PsbO that previously had been identified in this region.41,42 Radiolytic mapping was also used to oxidatively modify surface residues on PsbO that were in contact with water. Seventy-seven residues were identified. Interestingly, a large domain on the surface of PsbO was resistant to radiolytic modification, indicating that it was shielded from the bulk solvent. This domain appears to define buried regions of PsbO that are in contact with other, as yet unidentified, components of PS II.

### MATERIALS AND METHODS

#### PS II Membrane Isolation and Protein Cross-Linking. PS II membranes were isolated by the method of Berthold et al.43 from market spinach. Chl was determined by the method of Arnon.44 After isolation, the PS II membranes were suspended at a concentration of 2 mg of chl/mL in 50 mM Mes-NaOH (pH 6.0), 300 mM sucrose, 15 mM NaCl (SMN) buffer and frozen at −80 °C until they were used. Protein cross-linking was performed using the zero-length cross-linker EDC. In our experiments, PS II membranes were suspended at a chl concentration of 200 μg/mL in 25% glycerol, 10 mM MgCl₂, 5 mM CaCl₂, and 50 mM MES-NaOH (pH 6.0) buffer. The membranes were then treated with 6.25 mM EDC and 5 mM N-hydroxysulfosuccinimide for 2 h at room temperature in the dark. The reaction was quenched by bringing the reaction mixture to 100 mM ammonium bicarbonate and incubating the mixture for 20 min at room temperature. The membranes were harvested by centrifugation for 25 min at 38000g, and the final pellet was resuspended in 1.0 M NaCl in SMN buffer for 1 h at 4 °C to release the PsbP and PsbQ subunits. The PsbP- and PsbQ-depleted PS II membranes were washed with SMN and then treated with 1.0 M CaCl₂ in SMN buffer to release PsbO. After centrifugation to remove the PsbO-depleted membranes, the supernatant from the CaCl₂ wash (which contains PsbO) was dialyzed overnight against 10 mM Mes-NaOH (pH 6.0) using a 6−8 kDa cutoff membrane (Spectrum Laboratories, Inc.) and was concentrated by ultrafiltration using a 10 kDa cutoff membrane (Millipore Co.). Prior to electrophoresis, protein concentrations were determined using the BCA protein assay.45

#### Synchrotron Radiolysis. Synchrotron radiolysis was performed as described previously.46 Radiolysis was performed on the XLRM2 beamline at The J. Bennett Johnston, Sr. Center for Advanced Microstructures & Devices (CAMD) synchrotron. PS II membranes (200 μL at 2 mg of chl/mL) were exposed for various lengths of time at room temperature in a multichannel Plexiglas chamber. After exposure, the samples were immediately removed from the chamber and held on ice until being stored at −80 °C.

#### Electrophoresis and Protein Digestion. For the EDC cross-linking experiments, the protein samples were resolved on 12.5 to 20% LiDS−PAGE gradient gels66 using ammonium persulfate and tetramethylethylenediamine for polymerization. For the radiolytic experiments, however, the proteins were separated on 12.5 to 20% acrylamide LiDS−PAGE gradient gels using a nonoxidizing system in which the gels were polymerized with riboavin (in the presence of diphenyliodonium chloride and tolusene sulfonate) by exposure to UV light.47,48 The upper reservoir also contained thioglycolate. This was required, as standard PAGE polymerization conditions are known to introduce numerous protein oxidation artifacts.49 Earlier experiments indicated that PS II proteins resolved in the nonoxidizing gel system exhibited much lower levels of artifactual protein oxidation than when resolved using standard LiDS−PAGE (see Figure S1 of ref 48).

After electrophoresis, the gels were stained with Coomassie Blue and destained, and protein bands of interest were excised. The protein bands were processed for protease digestion (trypsin or trypsin with Lys-C) using standard protocols. After digestion, the proteolytic peptides were processed using a C18 ZipTip prior to mass analysis.

#### Mass Spectrometry. The proteolytic peptides were resolved using reversed-phase chromatography with mass spectrometry
being performed on a Thermo Scientific LTQ-FT instrument.\textsuperscript{48} This is a hybrid instrument consisting of a linear ion trap coupled to a Fourier transform ion cyclotron resonance mass spectrometer.

Identification and analysis of peptides containing cross-linked products or oxidative mass modifications were performed using the MassMatrix online search engine.\textsuperscript{50,51} A FASTA library containing PsbO, PsbP, and PsbQ proteins was searched, as was a decoy library containing the same proteins but with reversed amino acid sequences. For the identification of cross-linked products, peptides were selected if their P value was $\leq 0.0001$. In this study, as expected, no cross-linked products involving the PsbP and PsbQ proteins were identified. Additionally, cross-linked products were identified with StavroX version 3.4.12\textsuperscript{52} using the same FASTA library. This program reports a non-probabilistic score and a false discovery rate (FDR) for each identified cross-linked product. The FDR corresponded to 5%. Only putative cross-linked products that exhibited scores of $> 100$ (which was considered a “significant” score)\textsuperscript{49} and also exhibited scores $> 3$ times the FDR were considered in our study. In both MassMatrix and StavroX, a precursor ion precision of $\leq 5.0$ ppm was required.

For the radiolytic footprinting studies, MassMatrix was programmed to search for all of the possible oxidative modifications for the 18 modifiable and identifiable amino acids, excluding glycine and alanine.\textsuperscript{56,53} For the identification of oxidative modifications, a more stringent P value ($\leq 0.00001$) was used. We feel that given the smaller mass modifications introduced by oxidative labeling (2–48 au), the use of a more stringent P value was prudent. For both the protein cross-linking studies and the radiolytic modification experiments, the peptides were required to exhibit 0% hits to the decoy library for further consideration.

**Protein Modeling.** Clustal Omega\textsuperscript{54} was used to align the spinach and *Thermosynechococcus vulcanus* PsbO protein sequences. On the basis of this alignment, the spinach sequence was threaded using SWISS-MODEL\textsuperscript{59} with the structure of *T. vulcanus* PsbO [contained in Protein Data Bank (PDB) entry 3WU2, which is the update of PDB entry 3ARC\textsuperscript{60} as a template. Secondary structure prediction for higher-plant PsbO was performed using Genesilico Metaserver.\textsuperscript{58} The cross-linking distance constraints imposed by the 20 identified EDC cross-linked products were incorporated into the threaded structure using MODELLER.\textsuperscript{57} EDC generates an amide bond between primary amino groups and carboxylates that are in van der Waals contact.\textsuperscript{58,59} To model these interactions, a $C_{\alpha} – C_{\alpha}$ distance of 12.1 Å\textsuperscript{61} was used. This distance conservatively takes into account the length of the amino acid R groups and dynamic variations in the protein structure. The 10 lowest-energy models were analyzed by Ramachandran analysis using PROCHECK\textsuperscript{62} and RAMPAGE.\textsuperscript{63}

**Antibodies.** The antiserum against PsbO was produced using a synthesized peptide consisting of the N-terminal 27 amino acid residues of the mature spinach protein coupled to keyhole limpet hemocyanin. The antiserum was produced and isolated from rabbits after their immunization by standard protocols.\textsuperscript{64} The specificity of the antiserum was confirmed by its specific reaction with the antigenic peptide, isolated PsbO, PS II membrane preparations, and thylakoid membranes.

### RESULTS AND DISCUSSION

**Protein Cross-Linking.** Figure 1 illustrates the results obtained upon treatment of PS II membranes with the cross-linker EDC followed by LiDS–PAGE and immunoblotting.

![Figure 1](https://example.com/figure1.png)

**Figure 1.** EDC cross-linking of PsbO within the PSII complex. Protein cross-linking was performed as described in Materials and Methods. Illustrated are “Western” blots from the extracts of 1.0 M CaCl$_2$-washed PSII membranes (which were obtained after initially washing with 1.0 M NaCl) cross-linked with EDC. Panel A was stained with Coomassie blue, and panel B was probed with anti-PsbO. Intact PS II membranes are labeled PS II and are included for reference. The putative PsbO intramolecular cross-linked product is indicated by the cyan arrows and has an apparent molecular mass of 25 kDa. This protein is the subject of our cross-linking study.

After protein cross-linking, the PS II membranes were washed with NaCl to remove the PsbP and PsbQ proteins. These membranes were then washed with CaCl$_2$ to remove both unmodified PsbO and PsbO containing putative intramolecular cross-linked residues. This sample was then dialyzed and concentrated by ultrafiltration. As expected, in the absence of EDC, no cross-linked products were observed for PsbO. After cross-linking with EDC, a variety of cross-linked products containing PsbO were observed. Some of these remained associated with the PS II membranes after sequential washing with 1.0 M NaCl, which removes PsbP and PsbQ from the PS II membranes [no cross-linked products containing either PsbP or PsbQ were observed (data not shown)], and then washing with 1.0 M CaCl$_2$. These high-apparent molecular mass components represent intermolecular cross-linked products containing PsbO and intrinsic membrane protein components\textsuperscript{65,66} and are the object of continuing investigations. The CaCl$_2$ treatment removed un-cross-linked PsbO, which migrated at an apparent molecular mass of 29.6 kDa, and PsbO, which contained putative intramolecular cross-links, as indicated by more rapid migration via LiDS–PAGE, exhibiting an apparent molecular mass of 25 kDa. This protein band reacted only with anti-PsbO and not anti-PsbP or PsbQ (data not shown). This 25 kDa protein is the object of this work.

The observed 25 kDa band could be the result of either proteolysis or intrachain protein cross-linking. Subsequent tandem mass spectrometry indicated no proteolysis was evident as we obtained 100% coverage for PsbO, with complete coverage of the N- and C-termini. Consequently, the 25 kDa band appears to result from internal cross-linking with EDC.

In-gel proteolysis followed by tandem mass spectrometry allowed the identification of the cross-linked residues in this 25 kDa band (Table 1). Twenty intrachain cross-linked products were identified within PsbO. All of the cross-linked peptides identified exhibited low P values (the probability that the peptide match is a random occurrence) ranging from $1 \times 10^{-4}$ to
Table 1. EDC-Cross-Linked Residues Identified in PS II-Bound Spinach PsbO

<table>
<thead>
<tr>
<th>Residue</th>
<th>MassMatrix P value</th>
<th>Crystal structure distance (Å)</th>
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<tbody>
<tr>
<td>4K</td>
<td>5.0 × 10^{-5}</td>
<td>NA</td>
</tr>
<tr>
<td>6K</td>
<td>6.3 × 10^{-5}</td>
<td>NA</td>
</tr>
<tr>
<td>10K</td>
<td>5.0 × 10^{-7}</td>
<td>NA</td>
</tr>
</tbody>
</table>

**Structure of Higher-Plant PsbO.** Table 1 summarizes the cross-linked products identified using MassMatrix and StavroX. For the majority of the cross-linked products observed, analogous residue pairs are present in cyanobacterial PsbO. The Cα–Cα distances between these corresponding residue pairs in cyanobacterial PsbO are generally within 12.1 Å (Table 1); this was expected and highlights the overall structural similarity between the higher-plant and cyanobacterial PsbO proteins. Only two anomalously long distances were observed. In *T. vulcanus*, the residues that correspond to the 66K×234D cross-linked residues in spinach are residues 58K and 231E. In the crystal structure, the α-carbons of these residues are 16.1 Å apart. This might indicate a modest conformational difference of ~4 Å between the higher-plant and cyanobacterial proteins. The second anomaly is more difficult to explain. In *T. vulcanus*, the residues that correspond to the 60K×105K cross-linked residues in spinach are residues 60K and 105K. In the crystal structure, the α-carbons of these residues are 22.9 Å apart. This would indicate a rather large conformational (>10 Å) difference between the higher-plant and cyanobacterial proteins. Interestingly, these residues flank a domain on the PsbO protein that is resistant to radiolysis modification (see below). At this point in time, we cannot explain this anomaly.

The residues located at the N-terminus of the higher-plant protein (1E–10E) are not present in cyanobacterial PsbO. Three cross-linked residue pairs were present in this domain. Additionally, cross-linked residues were located in the vicinity of spinach residues 138P and 139E. The eight-residue insertion that forms the cyano loop in the *T. vulcanus* PsbO structure lies between 138P and 139E in the higher-plant protein. Finally, cross-linked residue pairs that lie near spinach residues 137D–138K that are absent in *T. vulcanus* PsbO were identified. This cross-linking information places constraints on the possible structures that can be assumed by higher-plant PsbO when it is bound to PS II.

Given these distance constraints provided by the observed cross-linked products (Table 1), molecular dynamics refinement can provide useful models for higher-plant PsbO, particularly with respect to its 10 N-terminal amino acid residues, which are not conserved in the cyanobacterial protein. MODELLER was used to provide the structures shown in Figure 3, which illustrates molecular dynamics structural refinements either in the absence (Figure 3A) or in the presence (Figure 3B) of the EDC-cross-linking distance constraints. In both panels, the 10 lowest-energy structures for PsbO are illustrated in different shades of blue and are aligned with the *T. vulcanus* protein (colored orange). Without inclusion of the distance constraints, the N-terminus is highly disordered. Inclusion of the distance constraints, however, yields a family of very similar structures, all of which exhibit nearly complete y- and b-ion (Figure 2C). This cross-linked species was additionally identified using MassMatrix and StavroX. The residues located at the N-terminus of the higher-plant protein (1E–10E) are not present in cyanobacterial PsbO. Three cross-linked residue pairs were present in this domain. Additionally, cross-linked residues were located in the vicinity of spinach residues 138P and 139E. The eight-residue insertion that forms the cyano loop in the *T. vulcanus* PsbO structure lies between 138P and 139E in the higher-plant protein. Finally, cross-linked residue pairs that lie near spinach residues 137D–138K that are absent in *T. vulcanus* PsbO were identified. This cross-linking information places constraints on the possible structures that can be assumed by higher-plant PsbO when it is bound to PS II.

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It should be noted that secondary structure analysis using the Genesilico MetaServer of the N-terminal domain of PsbO (1E–10E), which is not present in the cyanobacterial protein, indicates that the consensus secondary structure for this region
contains a short α-helix (\(\alpha\)-helix). This program utilizes seven independent secondary structure prediction algorithms. These algorithms uniformly predicted an α-helix at this location, and consequently, this was incorporated into the distance-constrained models.

Elucidation of the structure of the N-terminus of PsbO is critical for understanding the function of this component. Early reports indicated that the 16 N-terminal amino acid residues (1E−16Y) were required for binding to PS II and efficient oxygen evolution in higher plants. Subsequently, site-directed mutagenesis studies indicated that two binding determinants for PsbO are located in this N-terminal domain (Figure 4). One determinant (4K−10E) is found in higher plants (and green algae), and its deletion abolishes 50% of the oxygen evolving activity and results in the loss of 50% of the recombinant PsbO binding. The second binding determinant (15T−18E) is present in both higher plants and cyanobacteria; its deletion leads to a further loss of PsbO binding and very low oxygen evolution rates. These authors hypothesized that the deletion of 4K−10E prevented one of two copies of PsbO from efficiently binding to the photosystem. The deletion of 15T−18E led to the loss of a second copy. The locations of these two experimentally determined binding determinants are illustrated in Figure 4B. It should be noted that the biochemical evidence that suggests that two copies of the PsbO protein are present per PS II monomer in higher plants is quite strong (for in-depth reviews, see refs 13 and 26).

Radiolytic Footprinting of PsbO. Radiolytic footprinting allows the identification of amino acid residues that are exposed to bulk solvent. The •OH that is produced during radiolysis is extremely reactive and can modify 18 different residues that can be identified by mass spectrometry. Consequently, radiolytic footprinting is significantly more robust than other chemical modification techniques that can usually label only a few types of residues.

Table 2 and Figure 5 present the results from the radiolytic footprinting of the PsbO subunit in association with the PS II complex. Mass spectrometry coverage of the PsbO protein was
Upon integration over all of the irradiation time points (0, 4, 8, and 16 s), Table 2 identifies the residues and types of oxidative modifications observed. In Table 2, 77 oxidatively modified PsbO residues were identified. Using modern high-resolution and sensitivity mass spectrometers (FTICR and Orbitrap-class instruments), the oxidative modification of 18 amino acid residues can be detected and identified. Only glycine and alanine remain refractory to radiolytic analysis, because of their low reactivity.

Table 2. Oxidative Modifications of PsbO

<table>
<thead>
<tr>
<th>Residue</th>
<th>Type</th>
<th>Mass Shift (Da)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1-stcb</td>
<td>Y+do</td>
<td>+13.98</td>
</tr>
<tr>
<td>T1-stcb</td>
<td>D+gam</td>
<td>+31.99</td>
</tr>
<tr>
<td>E+ca/gam</td>
<td>17L+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>E+ca</td>
<td>16V+ca</td>
<td>+25.99</td>
</tr>
<tr>
<td>T1-stcb</td>
<td>14R+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>14R+go</td>
<td>13K+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>E+ca</td>
<td>12Q+ca</td>
<td>+14.99</td>
</tr>
<tr>
<td>E+ca</td>
<td>11I+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>S+stcb</td>
<td>14R+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>S+stcb</td>
<td>13K+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>13Q+ca</td>
<td>14R+go</td>
<td>+15.99</td>
</tr>
<tr>
<td>14R+go</td>
<td>13K+go</td>
<td>+15.99</td>
</tr>
</tbody>
</table>

“MassMatrix was used to identify the locations of oxidative modifications. In some instances, different types of oxidative modifications were observed for a given residue. Key: ca, carbonyl addition; +13.98 Da; do, double oxidation; +31.99 Da; go, general oxidation; +15.99 Da; gam, Glu/Asp decarboxylation; +30.01 Da; stcb, Ser/Thr carbonyl, −2.02 Da. It should be noted that while a total of 12 different types of oxidative modifications were incorporated into the MassMatrix searches, only these five types were actually observed.

100% upon integration over all of the irradiation time points (0, 4, 8, and 16 s). Table 2 identifies the residues and types of oxidative modifications observed. The individual time points are not shown; Table 2 and Figure 5 represent a unified set of all of the identified oxidatively modified residues observed in the experiment at all time points.
Figure 5. Radiolytic mapping of solvent-exposed domains on higher-plant PsbO. Seventy-seven oxidatively modified residues (blue spheres) were identified. These are listed in Table 2. The mass spectrometry coverage of PsbO was 100%. (A) View of PsbO from the surface of the PS II intrinsic proteins. Residues that were not oxidatively modified and associate with the intrinsic components of the photosystem are shown as red dotted spheres. (B) View of PsbO from the lumen looking down onto PS II. No oxidatively modified residues were identified in the large contiguous lumenal exposed domains labeled domain A (outlined in cyan) and domain B (outlined in orange). Residues that were not oxidatively modified and face the bulk solvent are shown as green dotted spheres. We speculate that these are shielded from oxidative modification by as yet unidentified PS II components. (C) Spinach PsbO shown within the context of cyanobacterial PS II. CP43 is colored green; CP47 is colored brown, and all other subunits are colored gray.

In higher-plant PS II dimer interface. These authors observed cross-linking between PsbO and CyanoQ that strongly supported their hypothesis. If PsbQ were positioned in an analogous location in higher-plant PsbO, suggesting that it lies at the periphery of PS II and is closely associated with CP43. In our view, this is highly unlikely and would require unprecedented conformational flexibility of the β-barrel of the protein. Additionally, it would require that the overall architecture of higher-plant PsbO be significantly different from that of the cyanobacterial protein. No evidence of this exists.

A second hypothesis is that the unmodified domains represent regions on the PsbO protein that are shielded by interaction with other PS II subunits. Several possibilities exist. The unmodified PsbO domains could be regions that interact with PsbP and PsbQ. Several investigators have examined the interaction of these subunits with the photosystem. In cyanobacteria, Liu et al. proposed a binding domain for CyanoQ at the lumenal PS II dimer interface. These authors observed cross-linking between PsbO and CyanoQ that strongly supported their hypothesis. If PsbQ were positioned in an analogous location in higher-plant PS II, this could explain the lack of radiolytic modification in domain A (Figure 5B,C), which lies near the PS II dimer interface. One such model was previously proposed for higher-plant PS II. It should be noted that other investigators have presented alternative models for the location of higher-plant PsbQ, suggesting that it lies at the periphery of PS II and is closely associated with CP43. In these models, PsbQ was suggested to be located in a position roughly similar to that of PsbQ in the red alga C. caldarium. If this positioning is correct for higher-plant PsbQ, then the observed absence of modifications in domain A (Figure 5B,C) cannot be explained. In any event, the presence of unmodified residues in domain B (Figure 5B,C) cannot be attributed to the presence of PsbQ in any of the models that have been presented previously.

It also appears very unlikely that PsbP could occupy domain B. In higher-plant PS II membranes, Ido et al. observed EDC cross-linking between PsbP and PsbE, indicating that these residues are in van der Waals contact. The closest approach between PsbE and domain B is >80 Å, which would appear...
to preclude the association of PsbP with domain B of PsbO (Figure 5B,C).

Other components have been reported to be associated with PS II membranes and core complexes. These include catalase and polyphenol oxidase, TL29, TL29, TL29, TL29, TL29 TL29, TL29, TL29 TL29, TL29, TL29 and a YCF39-like protein. It is possible that these (and perhaps other) components are associated with PS II membranes but have generally escaped detection, possibly because of the substoichiometric abundance or unusually poor protein staining. The presence of a substoichiometric component in association with domain B (Figure 5B,C), for instance, could reduce the substoichiometric abundance or unusually poor protein staining.

Within the PS II monomer is very strong, it must be stressed that no direct structural evidence of a second copy of PsbO has been forthcoming. At this time, we cannot distinguish between these various possibilities.

## CONCLUSIONS

In this work, we have provided new information concerning structural features of the PsbO protein from higher plants while it is in association with PS II membranes. The location and organization of the N-terminal 10-amino acid residue extension that is present in higher plants, but not in cyanobacteria, has been examined and structural models for this domain presented. This region appears to be instrumental in the binding of PsbO in higher plants. Additionally, radiolytic footprinting experiments indicate that large contiguous domains on the surface of PsbO are resistant to oxidative modification and, consequently, appear to be shielded from the bulk solvent by as yet unidentified PS II components. Ongoing investigations are underway in an attempt to identify the species responsible for this observation.

## NOTE ADDED IN PROOF

Recently, Wei et al. (2016) (Nature DOI: 10.1038/nature18020) have presented a cryo-EM structure of an inactive spinach PS II-LHCII supercomplex at 3.2 Å resolution. The N-terminus of the PsbO protein that is presented in this structure is highly congruent to the structure presented in this communication.

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**Notes**

The authors declare no competing financial interest.

**ABBREVIATIONS**

DOPE, discrete optimized protein energy; EDC, 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide; LiDS, lithium dodecyl sulfate; PAGE, polyacrylamide gel electrophoresis; PS I, Photosystem I; PS II, Photosystem II.

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